

A Picture is Worth a Thousand Words; Analyzing Clinical Data with Sequencing Data

Nalin Tikoo, Roche Sequencing Solutions, Pleasanton, USA
Hans-Peter Adams, Roche Sequencing Solutions, Potsdam, Germany
Priya Saradha, Experis IT Services US, LLC, New Jersey, USA
Aarthi Balasubramanyam, Roche Sequencing Solutions, Pleasanton, USA
Mathias Heibeck, Roche Sequencing Solutions, Potsdam, Germany

ABSTRACT

Advances in sequencing technologies have paved the path to better understanding cancer, hereditary diseases, infectious diseases, and prenatal diagnostics. This includes sequencing of germline DNA, tumor tissue DNA, and plasma DNA, i.e. liquid biopsy. Monitoring using plasma DNA compared to traditional imaging based methods has become routine clinical practice thus producing data in gigabyte to terabyte range and resulting in thousands of markers. Visualization and analysis of these data in the context of clinical data for diagnosis, prognosis, assessment of response to therapy, and outcome measures can be a challenge.

Aside from an introduction into the clinical and sequencing data, this paper will present various methods of visualization using SAS[®] software and R for giving informative views on the complex data. These visualizations serve the purpose of data review and analysis. Code and output snippets will be provided for subject profiling, custom survival analyses, bar charts and an RShiny application.

INTRODUCTION

According to the Precision Medicine Initiative¹, precision medicine is "an emerging approach for disease treatment and prevention that takes into account individual variability in genes, environment, and lifestyle for each person." Visualizing this massive amount of genomic data in conjunction with the clinical data is a challenge. The purpose of this paper is to showcase a few of these data visualizations that help us make sense of the data thus facilitating the development algorithms that further help with the patient care.

DATA

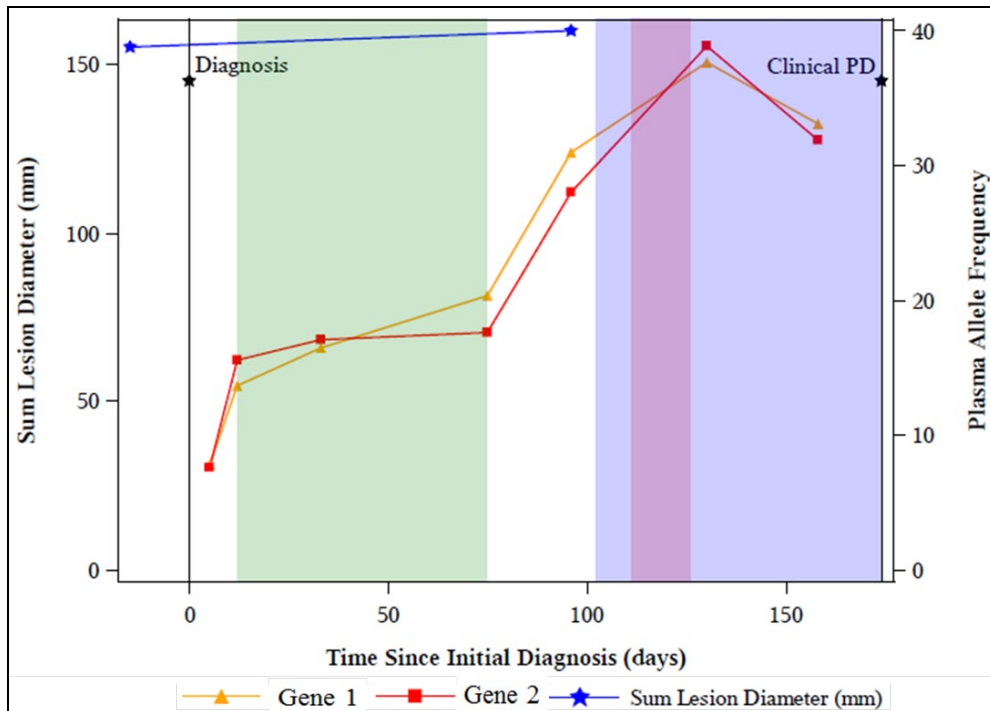
Clinical & Sequencing datasets were used for these visualizations. In terms of clinical data; demographics & baseline characteristics, Tissue & Plasma Sample collection, CT scans, treatment, treatment response measured by RECIST 1.1 and survival data were used. In terms of sequencing data, single nucleotide variant (SNV) variant call files (VCF) were used. The structure of this dataset is 1 record per patient per sample per gene variant.

PATIENT PROFILES – DISEASE MONITORING

DESCRIPTION

The objective of this visualization is to review the patient journey from diagnosis to clinical progression; represented by vertical reference lines on the two ends. Mutation data is plotted on the right y-axis, CT Scan data on the left y-axis and the time on study on the x-axis. The treatment information is represented using the colored bands.

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CODE SNIPPET

A macro was developed to loop through each subject's data; this was required to store subject specific values into macro variables to produce custom x-axis & y-axis value ranges & the lo and hi values for the treatment bands.

```
proc sgplot data=toplot_01. noautolegend;

    series x=monthtime y=af_plasma / group=gene markers name="afp" legendlabel="Plasma Allele Frequency"
    datalabel=af_plasma y2axis;
    y2axis label='Plasma Allele Frequency' min=0 max=%maxy2axisval.;

    series x=monthtime y=sumlesdiameter / lineattrs=(pattern=solid color=blue) markers markerattrs=(symbol=starFilled color=blue)
    name="slesd" legendlabel="Sum Lesion Diameter (mm)";
    yaxis label='Sum Lesion Diameter (mm)' min=0 max=%maxy1axisval.;

    scatter x=monthtime y=gediag / markerattrs=(symbol=StarFilled color=black) datalabel=gediagc datalabelattrs=(size=10);
    refline monthdiagdt / axis=x lineattrs=(color=black);

    scatter x=monthtime y=gepdc / markerattrs=(symbol=StarFilled color=black) datalabel=gepdc datalabelattrs=(size=10);
    refline monthpddt / axis=x lineattrs=(color=black);

    %if %sysevalf(&band1lo, ne .) %then %do;
        band y=x1 lower=%band1lo. upper=%band1hi. / fillattrs=(transparency=0.8 color=green);
    %end;

    %if %sysevalf(&band2lo, ne .) %then %do;
        band y=x2 lower=%band2lo. upper=%band2hi. / fillattrs=(transparency=0.8 color=red);
    %end;

    %if %sysevalf(&band3lo, ne .) %then %do;
        band y=x3 lower=%band3lo. upper=%band3hi. / fillattrs=(transparency=0.8 color=blue);
    %end;

    xaxis min=%minxaxisval. max=%maxxaxisval.;
    keylegend "afp" "slesd" / across=3 autoitemsize;
run;
```

1. Right y-axis – Plasma Allele Frequency (%)
2. Left y-axis – Sum of Lesion Diameters (mm)
3. Vertical reference lines
 - a. First reference line for Diagnosis Date
 - b. Second reference line for Date of Disease Progression
4. Treatment bands – lower & upper bounds of the band are the number of days since diagnosis date (Day 0) to the start and end of each treatment respectively.
 - a. Green band for chemotherapy
 - b. Red band for radiation therapy

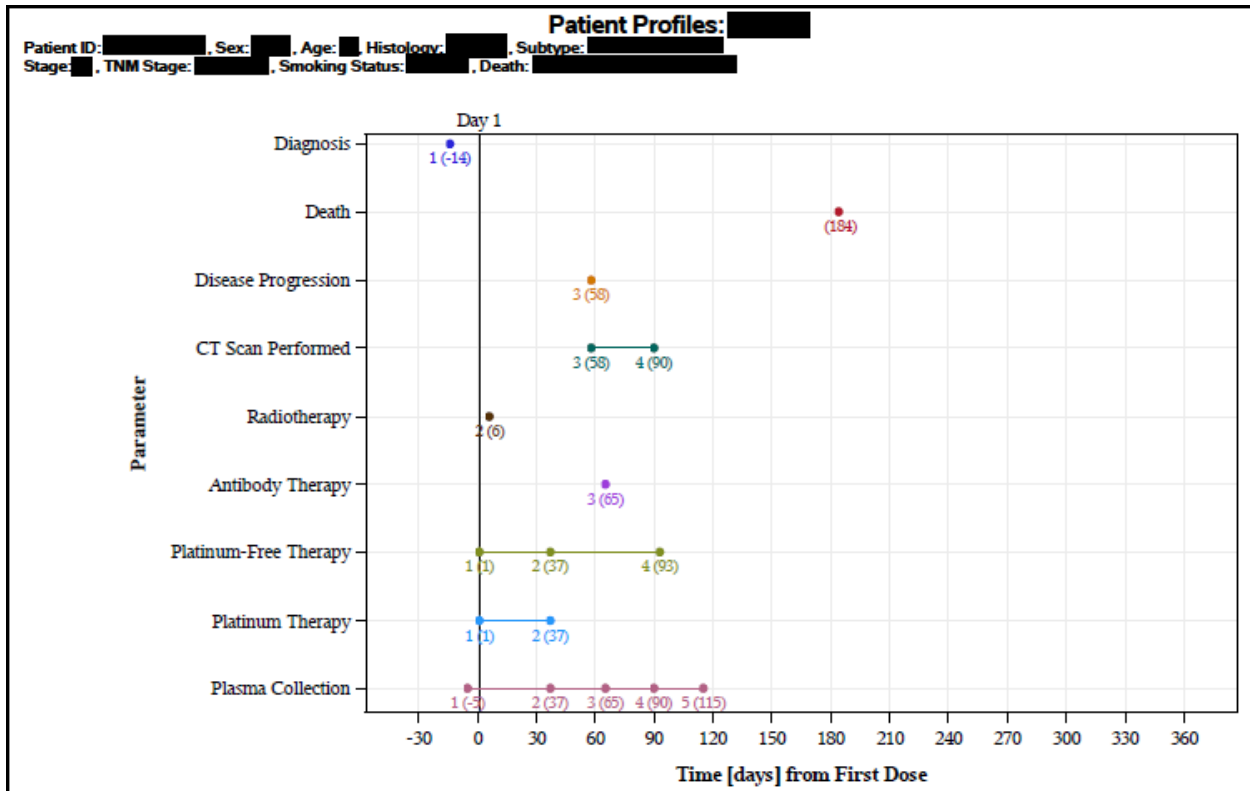
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- c. Blue band for targeted therapy

PATIENT PROFILES – DATA REVIEW FOR COMPLETENESS AND ACCURACY

DESCRIPTION

The objective of this visualization is to provide a visual of the expected events over time. This aids data management and clinical operations colleagues to query the implausible data points while helping clinicians and statisticians with developing imputation algorithms.



CODE SNIPPET

The key to this visualization is generating the dataset. The structure of the dataset needs to be 1 record per event with the number of days from first dose date to that event.

```
proc sgplot data=toplot_final_61. noautolegend nocycleattrs;
  refline 1 / axis=x lineattrs=(thickness=1 color=black) label="Day 1";

  scatter x=aval y=param/markerattrs=(size=7px symbol=circlefilled) group=param
    datalabel=avisit
    datalabelpos=bottom
    colormodel=(CX3288BD CX99D594 CXE6F598 CXFEE08B CXFC8D59 CXD53E4F);

  series x=aval y=param/group=param markers markerattrs=(symbol=circlefilled size=7px)
    lineattrs=(pattern=solid);

  axis grid values=(-30 to 375 by 30);
  yaxis grid values=("Plasma Collection" "Platinum Therapy" "Platinum-Free Therapy" "Antibody Therapy"
    "Radiotherapy" "CT Scan Performed" "Disease Progression" "Death" "Diagnosis");
run;
```

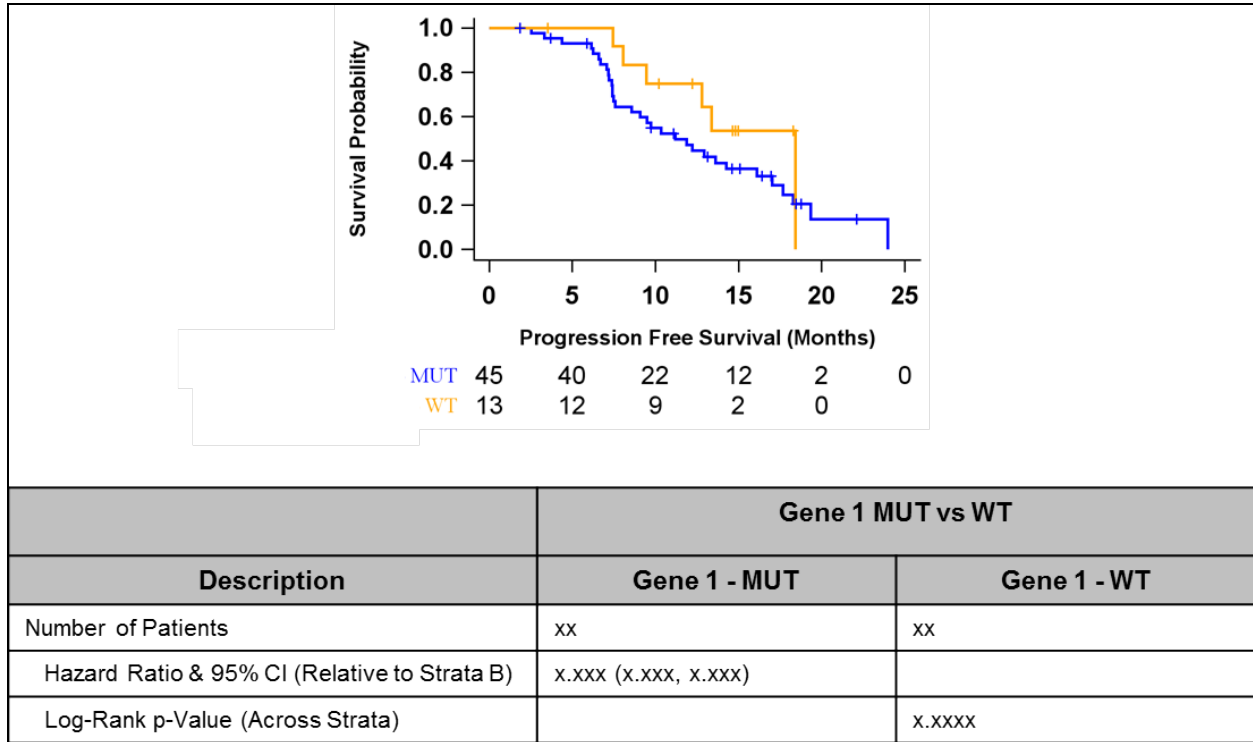
1. DATALABEL option is used to annotate the visit number and the number of days since first dose date.
2. DATALABELPOS option is used to provide the location of the annotation.

KAPLAN-MEIER CURVES

DESCRIPTION

The objective of this visualization is to present the K-M curves including relevant statistics on one PowerPoint slide. The K-M curves were customized to very specific specifications to meet publication requirements.

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CODE SNIPPET

Custom K-M Curve

The user guide² provides details on the usage of the COMPILESURVIVALTEMPLATES & PROVIDESURVIVALMACROS macros provided by SAS for producing highly customized K-M curves. The code snippets below highlight a few key customizations that we applied.

```

*** Custom Line Pattern & Colors ***;
%let GraphOpts = attrpriority=none DataContrastColors=(blue orange grey green)
                  DataLinePatterns=(solid)
                  DataColors=(blue orange grey green);

*** Custom Fonts ***;
%let yOptions = label="Survival Probability" shortlabel="Survival" labelattrs=(size=
                  tickvalueattrs=(size=12pt weight=bold f
                  linearopts=(viewmin=0 viewmax=1);
%let xOptions = shortlabel=XNAME offsetmin=.05 linearopts=(viewmax=MAXTIME tickvalue
                  labelattrs=(size=10pt weight=bold fam
                  tickvalueattrs=(size=12pt weight=bold

*** At Risk Table Numbers Color Size & Font Family ***;
%let AtRiskOpts = valueattrs=(size=12pt color=black family='Arial');

*** p-value table location ***;
%let InsetOpts = ;

*** Group Legend location ***;
%let LegendOpts = ;

*** Step Curve Thickness ***;
%let StepOpts = lineattrs=(thickness=2);

```

1. Customizing the colors used for the strata values

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2. Customizing the text attributes for the At Risk table
3. Customizing the thickness of the step curve

One thing worth noting is that when xOptions and yOptions macro variables are used to customize xAxis and yAxis values, the default values for these macro variables need to be included in the custom call.

Printing on one slide

The paper³ provides details on the usage of ODS POWERPOINT and ODS LAYOUT in conjunction. The ODS statements provide a variety of powerful features; some of the features that we used are described below.

```
options nodate nonumber orientation=landscape;
ods powerpoint file="xx\xx\xx\xx\xx.pptx" nogtitle
                                nogfootnote
                                style=rsspptstyle
                                dpi=300;
ods graphics on / noborder width=3.5in height=2.5in;

ods layout gridded rows=2 row_gutter=5pct; 1a
ods region;                                1b
*** K-M Curve Code ***;                    1c
ods region;                                1d
*** Survival Statistics Table Code ***;
ods layout end;

data _null_;
  dcl odsout obj(); 2
run;

ods layout gridded rows=2 row_gutter=5pct;
ods region;
*** K-M Curve Code ***;
ods region;
*** Survival Statistics Table Code ***;
ods layout end;

ods powerpoint close;
```

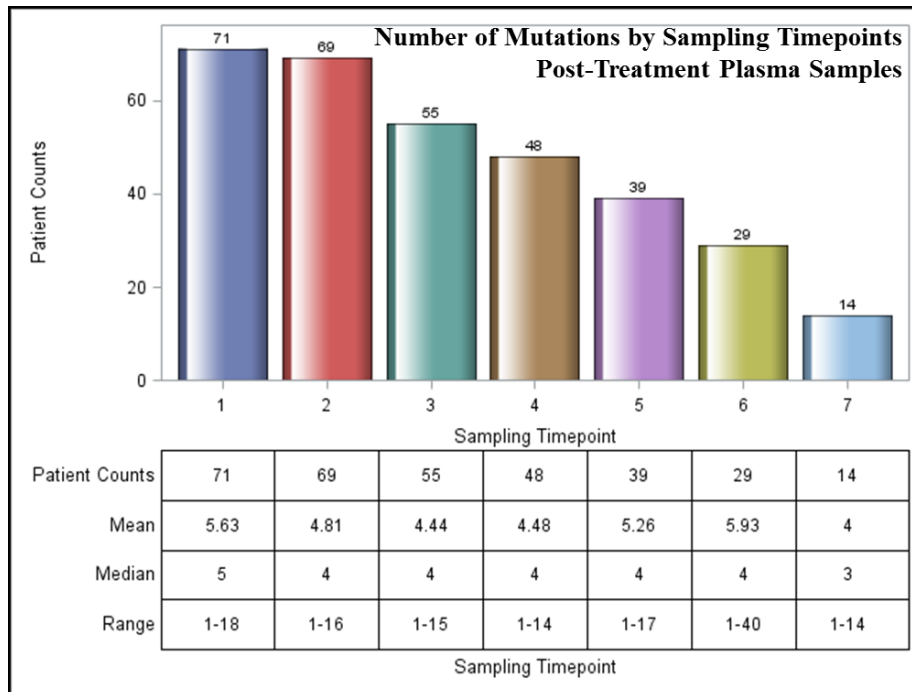
1. Code for Slide#1
 - a. The ODS LAYOUT GRIDDED statement starts the layout container. The ROWS= option is set to 2 so that the output is stacked together.
 - b. The ODS REGION statement begins the region container that is to hold the K-M curve.
 - c. The ODS REGION statement begins the region container that is to hold the statistics table.
 - d. The ODS LAYOUT END statement ends the gridded layout.
2. Code for triggering a new slide.
3. Code for Slide#2

BAR CHARTS

DESCRIPTION

The objective of this visualization is to understand the mutation profile for a study over time as seen in the visual below. It helps to easily and quickly know if the number of mutations per sample is within the expected range given the parameters for the study.

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CODE SNIPPET

The TEMPLATE procedure was used to produce a custom template to produce the table below the bar chart. This template was then used in the SGRENDER procedure to generate the final output.

```
proc template;
  define statgraph barblock;
    begingraph;
      layout lattice / rowweights=(.65 .35); 1
      layout overlay;
        barchart x=visit y=n / group=visit 2
          barlabel=true dataskin=gloss;
      endlayout;
      layout overlay / xaxisopts=(type=discrete display=(label))
        walldisplay=none;
      blockplot x=visit block=stats / class=desc 3
        display=(outline values label)
        repeatedvalues=true;
      endlayout;
    endlayout;
  endgraph;
end;
run;
```

1. **layout lattice** statement is used to create 2 rows; first row occupying 65% of the space displays the bar chart and the second row occupying 35% of the space displays the summary statistics table.
2. **barchart** statement is used to create the bar chart.
3. **blockplot** statement is used to create the summary statistics table below the bar chart.

RSHINY APPLICATION

DESCRIPTION

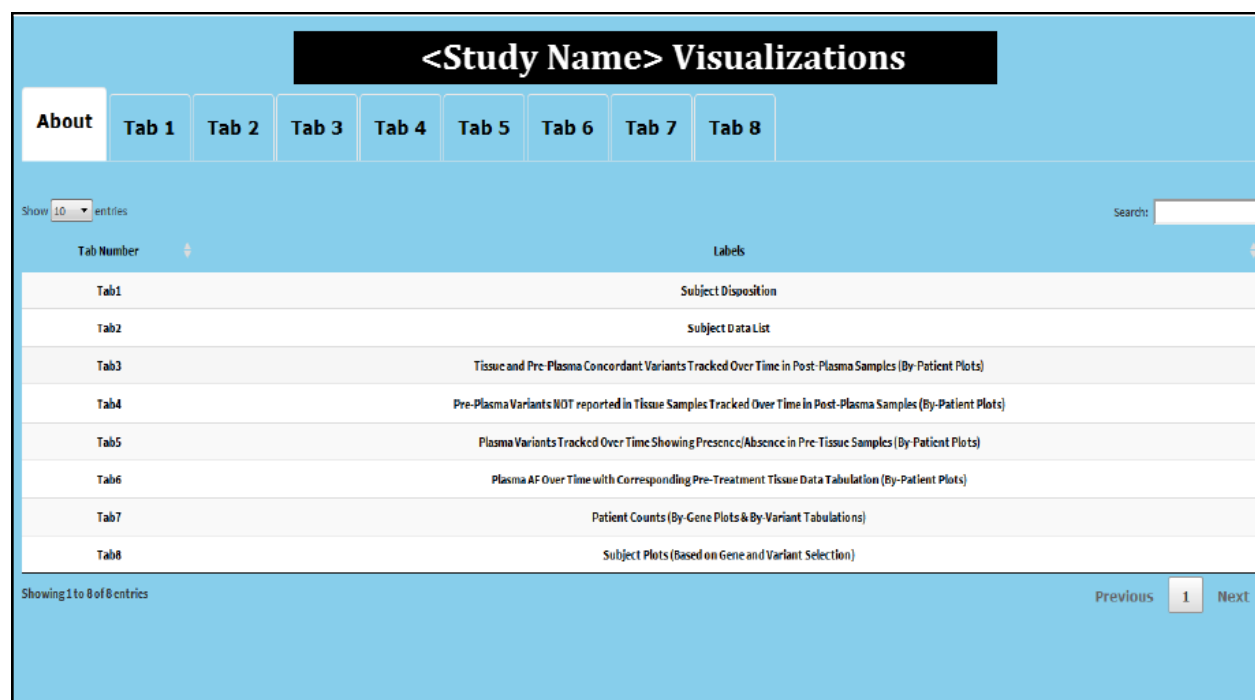
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The objective of this application was to provide the users with different visualizations to gain insights into the gene mutations for cancer patients with the help of interactive graphs and tabulations. The input data is a combination of genomic data generated by next-generation-sequencing and clinical endpoint data.

Shiny is an R package that was used to build the application. This package allows building interactive web applications straight from R. In addition to Shiny; the following R packages were used:

Package Name	Purpose
SHINY	Application user interface and server control
GGPLOT2	Dependency package for PLOTLY
PLOTLY	Interactive plots
RCOLORBREWER	Additional color components
GRDEVICES	Base color components
DT	Interactive data tables
PLYR	Required for data manipulation in R script being invoked
DPLYR	Required for data manipulation in R script being invoked
RESHAPE2	Required for data manipulation in R script being invoked
SAS7BDAT	Reading SAS files (optional)

The home page for the app lists the visualizations which the user can then navigate to by clicking on the corresponding tab number at the top.



<Study Name> Visualizations

About Tab 1 Tab 2 Tab 3 Tab 4 Tab 5 Tab 6 Tab 7 Tab 8

Show 10 entries

Search:

Tab Number	Labels
Tab1	Subject Disposition
Tab2	Subject Data List
Tab3	Tissue and Pre-Plasma Concordant Variants Tracked Over Time in Post-Plasma Samples (By-Patient Plots)
Tab4	Pre-Plasma Variants NOT reported in Tissue Samples Tracked Over Time in Post-Plasma Samples (By-Patient Plots)
Tab5	Plasma Variants Tracked Over Time Showing Presence/Absence in Pre-Tissue Samples (By-Patient Plots)
Tab6	Plasma AF Over Time with Corresponding Pre-Treatment Tissue Data Tabulation (By-Patient Plots)
Tab7	Patient Counts (By-Gene Plots & By-Variant Tabulations)
Tab8	Subject Plots (Based on Gene and Variant Selection)

Showing 1 to 8 of 8 entries

Previous 1 Next

TAB 1 – SUBJECT DISPOSITION

The purpose of this table is to provide a summary of the sample & subject inventory for the study.

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About	Tab 1	Tab 2	Tab 3	Tab 4	Tab 5	Tab 6	Tab 7	Tab 8
Subjects in study: [REDACTED] Sequenced and Analyzable Samples*: Pre-Tissue: [REDACTED] Pre-Plasma: [REDACTED] Post-Plasma: [REDACTED] Subjects with both Pre-Tissue and Pre-Plasma samples: [REDACTED] Only Pre-Tissue: [REDACTED] Only Pre-Plasma: [REDACTED] Subjects with both Pre-Plasma and Post-Plasma samples: [REDACTED] Only Pre-Plasma: [REDACTED] Only Post-Plasma: [REDACTED] *Sequenced & Analyzable: Samples used for variant calling								

TAB 2 – SUBJECT DATA LISTING

The purpose of this table is to provide a detailed data dump for all the subjects in the study.

Key Features:

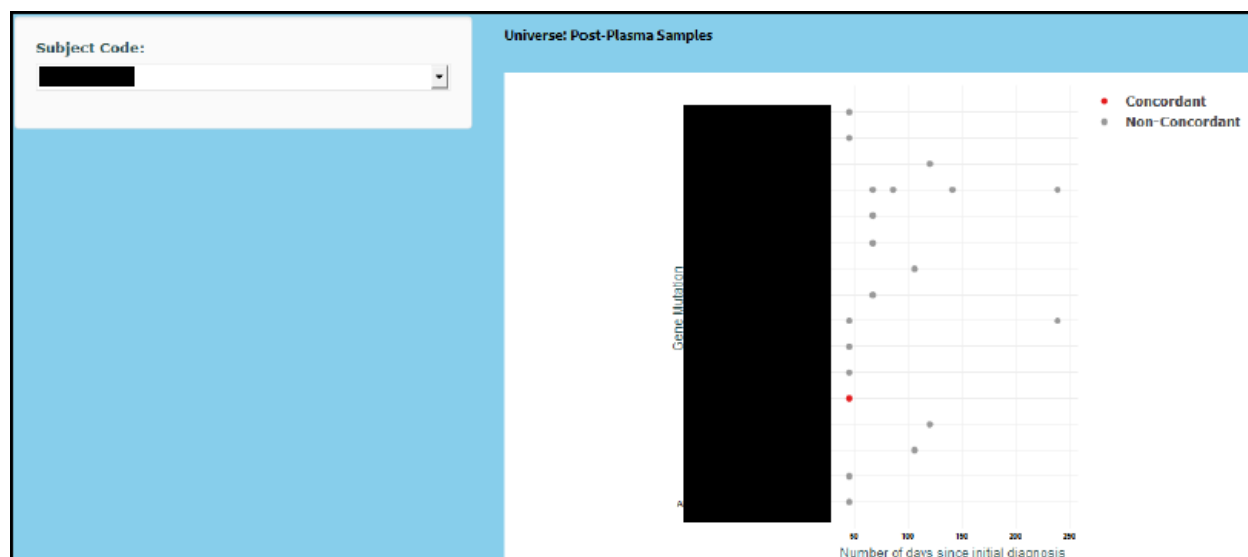
- The “Subject Code” selector on the left allows the user to review data from a selected patient(s) at a time.
- The text boxes at the top of each column in the table allow for selecting specific values in that column. As an example; for the GENE column, the user could select a particular Gene of interest to look at across the entire population.
- There is also an option to download the data in an Excel file format.

About	Tab 1	Tab 2	Tab 3	Tab 4	Tab 5	Tab 6	Tab 7	Tab 8																																																															
Subject Code: [REDACTED] Show 10 entries Search: <table> <thead> <tr> <th>Subject ID</th> <th>Sample Type</th> <th>No. of days since diagnosis</th> <th>TIMEPOINT</th> <th>GENE</th> <th>CDS</th> <th>AA</th> <th>AF</th> <th>SNVAFMEAN</th> </tr> </thead> <tbody> <tr> <td>[REDACTED]</td> <td>Tumor</td> <td>2</td> <td>Pre</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> </tr> <tr> <td>[REDACTED]</td> <td>cDNA</td> <td>23</td> <td>Pre</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> </tr> <tr> <td>[REDACTED]</td> <td>cDNA</td> <td>23</td> <td>Pre</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> </tr> <tr> <td>[REDACTED]</td> <td>cDNA</td> <td>23</td> <td>Pre</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> </tr> <tr> <td>[REDACTED]</td> <td>cDNA</td> <td>23</td> <td>Pre</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> </tr> <tr> <td>[REDACTED]</td> <td>cDNA</td> <td>23</td> <td>Pre</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> <td>[REDACTED]</td> </tr> </tbody> </table>									Subject ID	Sample Type	No. of days since diagnosis	TIMEPOINT	GENE	CDS	AA	AF	SNVAFMEAN	[REDACTED]	Tumor	2	Pre	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	cDNA	23	Pre	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	cDNA	23	Pre	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	cDNA	23	Pre	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	cDNA	23	Pre	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	cDNA	23	Pre	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]	[REDACTED]
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TAB 3 – CONCORDANT VARIANTS FROM PRE-TREATMENT TISSUE & PLASMA SAMPLES TRACKED OVER TIME IN POST TREATMENT PLASMA SAMPLES

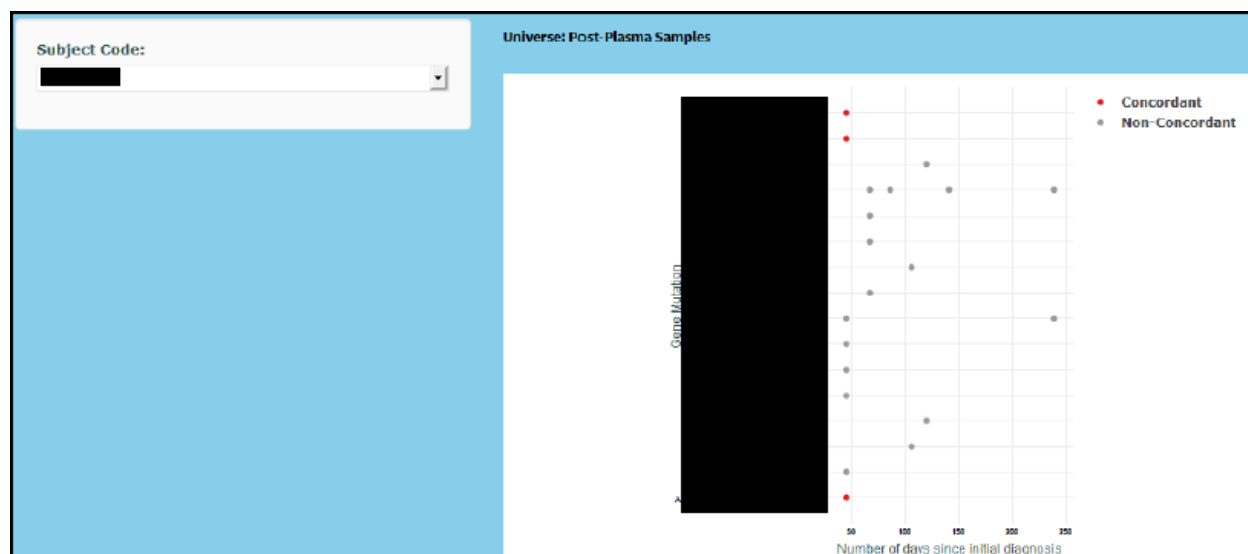
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The purpose of this visualization is to review & compare concordant gene mutations seen in pre-treatment samples vs post-treatment plasma samples.



TAB 4 – DISCORDANT PLASMA VARIANTS FROM PRE-TREATMENT SAMPLES TRACKED OVER TIME IN POST TREATMENT PLASMA SAMPLES

The purpose of this visualization is to review & compare discordant gene mutations seen in plasma samples; both pre & post treatment.



Only a handful of visualizations have been described in detail here. As can be seen in the list on the home page various other visualizations were developed in collaboration with stakeholders.

CONCLUSION

With the increasing volume of data being generated, data visualization has become an integral part of data analysis more than ever before. These visualizations aid in better understanding the data before adopting traditional statistical models for further analysis. The visualizations presented here are an example of that exercise.

From a programming perspective; an important aspect is to understand "fit-for-purpose" programming package. In addition to SAS® software & R described in this paper, Spotfire is another visualization package that can be used. All these are great packages & the key to figure out which one works best for your particular visualization needs.

REFERENCES

1. Understanding Genetics

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 - a. <http://support.sas.com/kb/49/697.html>
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 - a. <http://support.sas.com/resources/papers/proceedings16/SAS5443-2016.pdf>
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 - a. <http://documentation.sas.com/?docsetId=grstatproc&docsetTarget=n1t2ihc734azuvn17021ri2cl057.htm&docsetVersion=9.4&locale=en>
6. Shiny
 - a. <https://shiny.rstudio.com/>

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CONTACT INFORMATION

Your comments and questions are valued and encouraged. Contact the author at:

Author Name: Nalin Tikoo
Company: Roche Sequencing Solutions
Address: 4300 Hacienda Drive
City / Postcode: Pleasanton, 94588
Work Phone: 925-353-5384
Email: nalin.tikoo@roche.com
Web: <http://sequencing.roche.com/en.html>

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