

Nonclinical Study Data Specifications and CRO-Sponsor Workflows for FDA SEND Compliance

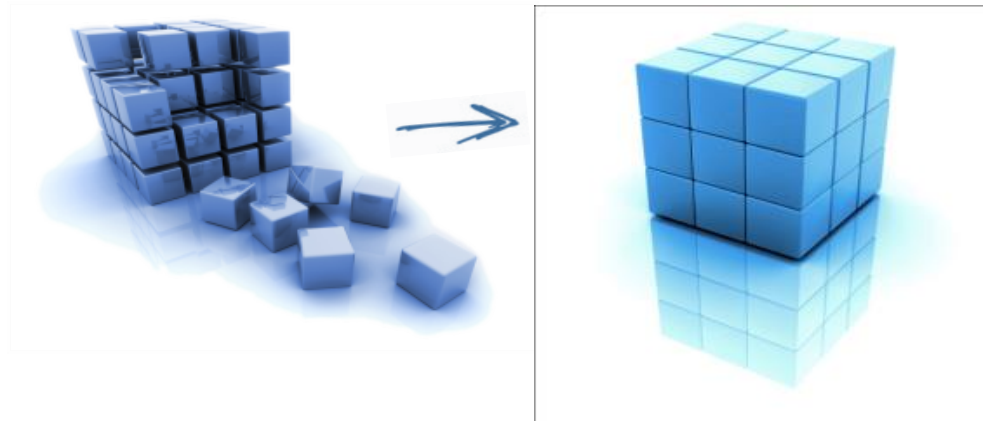
Kristi Johnson, PointCross Life Sciences
Michael Dorato, Smithers Avanza



Single Day Event
Foster City, CA
December 2, 2015

December 2, 2015

www.pointcrosslifesciences.com



Agenda

Topic	Presenter
Submission and Review Workflows at the FDA	Kristi Johnson
Current State of SEND in the Industry	Kristi Johnson
Challenges in Preparing SEND Datasets and CRO-Sponsor Workflows to Improve Data Quality	Kristi Johnson
Perspectives from a CRO	Michael Dorato
Summary, Q&A	All

Data flow at the FDA from submission to review

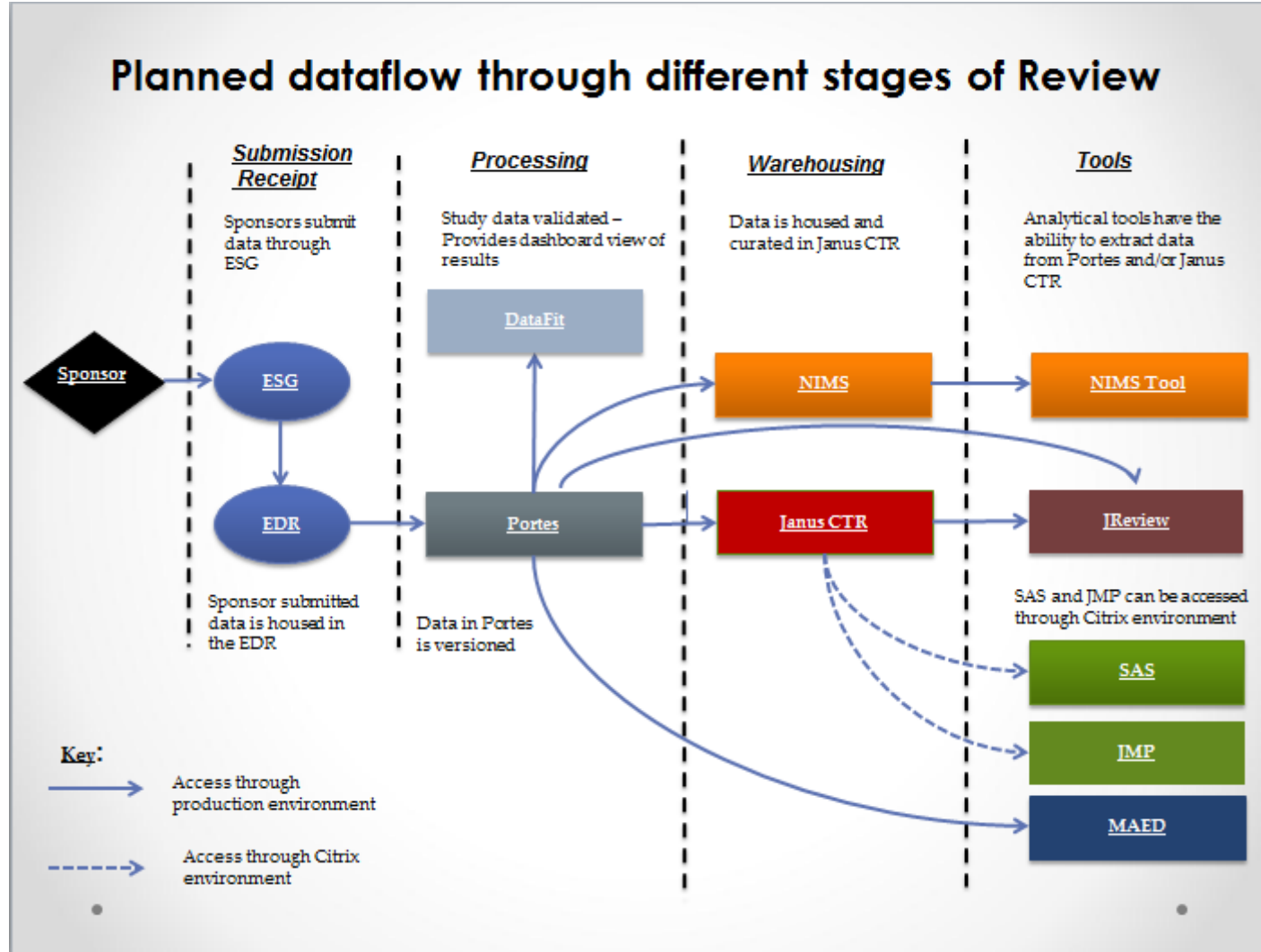


Diagram courtesy of Booz Allen Hamilton on behalf of FDA CDER

SEND Study Data Review Workflows at FDA CDER

- ✓ SEND datasets packaged in the M4 folder of eCTD submissions are received
- ✓ The Portes software versions the available studies in the submissions
- ✓ NIMS picks up new studies from Portes through automated workflows
- ✓ All SEND files are validated against CDISC and FDA validation rules
 - Dashboard populated
 - Validation reports, Define.xml and SDRG files are made available
 - Studies with “no validation errors” are loaded and ready for pharm/tox review
 - Studies with “validation errors” are not loaded; returned to submitter
- ✓ Reviewers access study data through tools like the ToxVision SEND data browser
 - Review tabulated or graphical views of findings and signals
 - Create clubbed groups or generate “sponsor groups” with exclusions
 - Check against Study Report to verify signals of interest

Potential Data Problems & Consequences to Sponsors

- ✓ Validation checks only assess the data format against rules
 - Failure means the data will not be loaded for review
 - Success can still mean issues at other stages
- ✓ Define.xml and study data files do not match (missing, or unexpected domains)
- ✓ If reviewers discover variance in summary data between re-constituted SEND trial sets to sponsor defined groups in the study report – ***Raises questions about the submission!***
- ✓ If reviewers see a signal in Lab-test data or incidence counts not visible in the study report individual subject results – ***Raises questions about the submission!***
 - Why are the individual data not the same in SEND and Study Report?
- ✓ If there are insufficient domains or data for the purposes of review and if the data is available in the study report – ***Raises possible request for additional data to be submitted***

Current Industry Situation regarding SEND

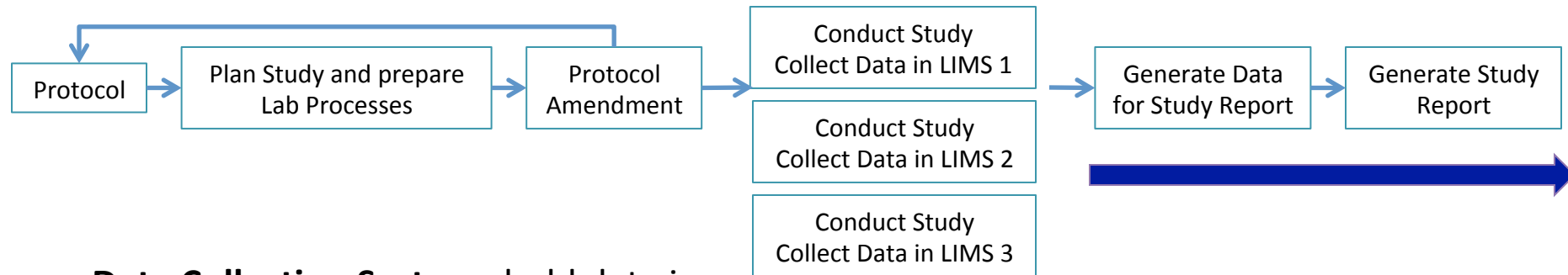
1. Increasing evidence that validating a SEND dataset for conformance to the standard is necessary *but simply not sufficient* to be SEND ready
 - SEND datasets that do not match tabulated data in the reports OR that are missing chunks of data will appear ***untrustworthy*** to FDA reviewers
2. The FDA's test submission process only provides a validator report which indicates if there are errors:
 - This is an *extremely low bar* to judge CRO and sponsor readiness
 - Only a real submission to the FDA will trigger regulatory review of the SEND data and the accompanying study report
 - **Sponsors, do you want to wait until you have a real submission to verify your providers? *Delays due to rejected data or questions about data integrity?***

Sponsor and CRO Challenges with SEND Preparation

A complete SEND submission should include *standardized SEND datasets* plus the *define file* and a *Study Data Reviewer's Guide*

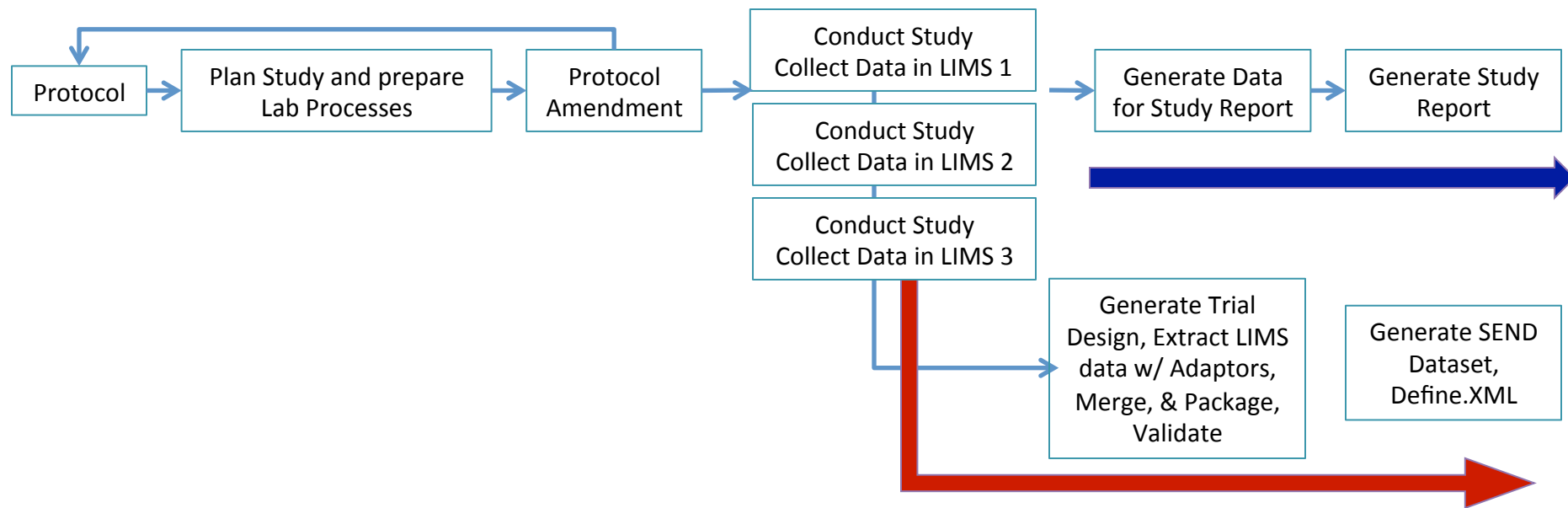
- ✓ Studies include data from various data providers/systems - must **harmonize and assemble** these different data streams into a single package
- ✓ Providing SEND datasets **technically conformant** with the CDISC standard, controlled terminologies and FDA specific validation rules
- ✓ Ensuring that the SEND data for every study is *fit for FDA review*:
 - **Consistent** with the study report
 - **Complete** with respect to all of the reportable data

Current Study Reporting Processes



- **Data Collection Systems** hold data in proprietary data models not easily accessed other than through system reports
- Keeping up with the collection system versions, data model changes and terminology changes is complex
- LIMS data are held in a GLP store
- LIMS data is transformed into preferred terms and study designs for Study Data publication by the Principal Investigator – see *the process flow*

Pitfalls of separate SEND & study reporting processes



- SEND datasets are generated by a parallel process possibly by different people using Trial Designs needed for SEND. The data is reported using “preferred” terminology of the toxicologists. *These do not match the study report’s dose groupings - see* ➡
- Study Reports are signed by the Study Director. The SEND Dataset must also be signed by the Sponsor or preparer.
- Because the two are constructed using disparate processes, they need NOT be consistent - to prove that they have the **right quality, consistency and completeness** is difficult, expensive, and time consuming

Examples of Data Conformance, Consistency and Completeness Issues in SEND Datasets & Submissions

Failure to comply with the SEND Standard

	STUDYID	DOMAIN	USUBJID	MASEQ	MATESTCD	MATEST	MAORRES	MASTRESC	MASPEC	MAANTREG	MALAT	MADTC	MADY
1	ABC123	MA	1	1	GROSPATH	Gross Pathological Examination	Enlargement	Enlargement	LYMPH NODE	BILATERAL		9/21/2015	48
2	ABC123	MA	2	1	GROSPATH	Gross Pathological Examination	Discoloration, Pale	Pale	SPLEEN			9/21/2015	48
3	ABC123	MA	3	1	GROSPATH	Gross Pathological Examination	Discoloration, Pale	Pale	LIVER			9/21/2015	48
4	ABC123	MA	4	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
5	ABC123	MA	5	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
6	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
7	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
8	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
9	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
10	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
11	ABC123	MA	11	1	GROSPATH	Gross Pathological Examination	Not Collected	Not Collected	SKIN			9/21/2015	48
12	ABC123	MA	11	2	GROSPATH	Gross Pathological Examination	Discoloration, Pale	Pale	LIVER			9/21/2015	48
13	ABC123	MA	11	3	GROSPATH	Gross Pathological Examination	Enlargement	Enlargement	LIVER			9/21/2015	48
14	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
15	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
16	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
17	ABC123	MA				Pathological Examination	Discoloration, Pale	Pale	KIDNEY	BILATERAL		9/21/2015	48
18	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
19	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
20	ABC123	MA				Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
21	ABC123	MA	19	1	GROSPATH	Gross Pathological Examination	Nodule		SKIN			9/21/2015	48
22	ABC123	MA	20	1	GROSPATH	Gross Pathological Examination	Nodule		SKIN			9/21/2015	48
23	ABC123	MA	21	1	GROSPATH	Gross Pathological Examination	Normal		ALL TISSUES			9/21/2015	48
24	ABC123	MA	21	2	GROSPATH	Gross Pathological Examination	Normal		ALL TISSUES			9/21/2015	48
25	ABC123	MA	21	3	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
26	ABC123	MA	21	4	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
27	ABC123	MA	22	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
28	ABC123	MA	23	1	GROSPATH	Gross Pathological Examination	Enlargement	Enlargement	Lymph Nodes	BILATERAL		9/21/2015	48
29	ABC123	MA	24	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
30	ABC123	MA	25	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
31	ABC123	MA	26	1	GROSPATH	Gross Pathological Examination		NORMAL	ALL TISSUES			9/21/2015	48
32	ABC123	MA	27	1	GROSPATH	Gross Pathological Examination		NORMAL	ALL TISSUES			9/21/2015	48
33	ABC123	MA	28	1	GROSPATH	Gross Pathological Examination		NORMAL	ALL TISSUES			9/21/2015	48
34	ABC123	MA	29	1	GROSPATH	Gross Pathological Examination		NORMAL	ALL TISSUES			9/21/2015	48
35	ABC123	MA	30	1	GROSPATH	Gross Pathological Examination		Nodule	SKIN			9/21/2015	48
36	ABC123	MA	31	1	GROSPATH	Gross Pathological Examination		Nodule	SKIN			9/21/2015	48
37	ABC123	MA	32	1	GROSPATH	Gross Pathological Examination	Enlargement	Enlargement	KIDNEY	BILATERAL		9/21/2015	48
38	ABC123	MA	33	1	GROSPATH	Gross Pathological Examination	Discoloration, Pale	Pale	KIDNEY	BILATERAL		9/21/2015	48
39	ABC123	MA	34	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48
40	ABC123	MA	35	1	GROSPATH	Gross Pathological Examination	Normal	NORMAL	ALL TISSUES			9/21/2015	48

Modifier has been incorrectly populated as the base finding

Specimen that was not collected has not had status populated as "NOT DONE"

Specimen modifier has been incorrectly classified

Variable populated with term not present on appropriate CT list

Consequences of Data Quality Issues

- ✓ Data consistency issues between the SEND data and study report may cause reviewers to question the *trustworthiness* of the data
- ✓ The SEND dataset lacks data present in the study report. This can cause reviewers to question why those data were excluded, and *delay the review process*
- ✓ Specimen information missing for Clinical Chemistry measurements and Gross Pathology. Datasets are *difficult to review and interpret* causing further delays
- ✓ Inability to load dataset due to syntax errors within Define.XML – *affects validation and automated loading*

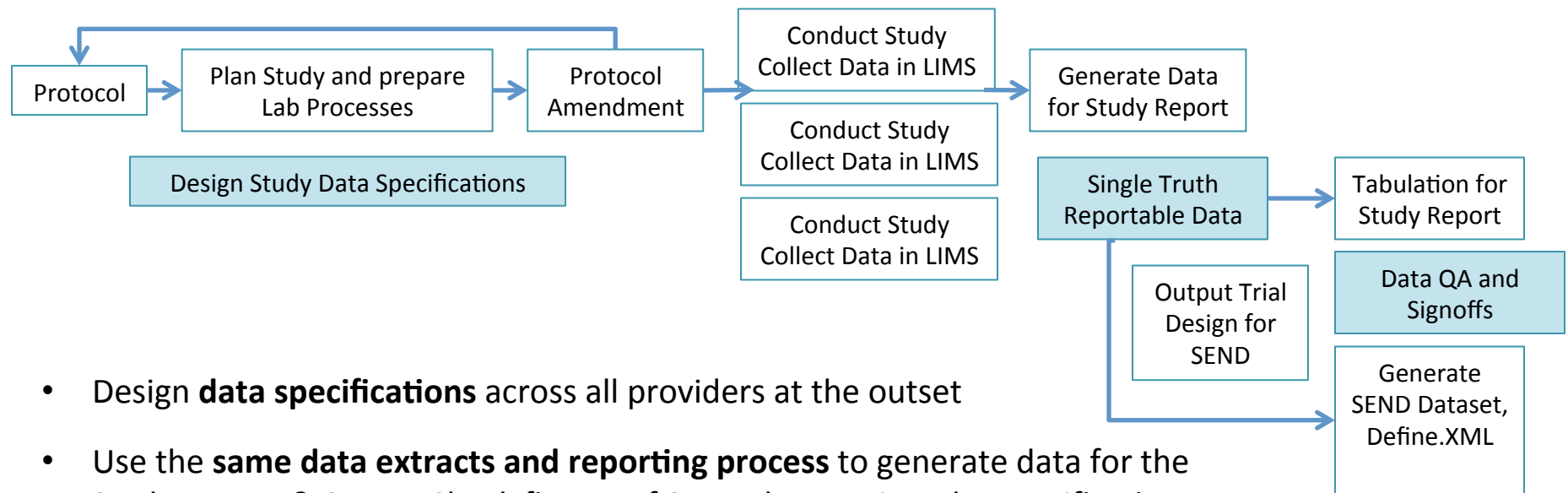
How to address data quality, consistency and completeness issues in SEND datasets

CRO-Sponsor workflows to improve data quality

- 1. Study Data Specifications:** Plan for SEND and specify expectations with each provider against which progress and performance can be measured and controlled
- 2. Single Truth Data Resource:** Use the same data resource in a “single truth” repository for both study reporting and automation of SEND transformation workflows
- 3. QA/QC tools and governance processes:** for a high quality product with reduced costs

Specify, Generate and Review Reportable Data

Data for study reporting and SEND in a common repository



- Design **data specifications** across all providers at the outset
- Use the **same data extracts and reporting process** to generate data for the Study Report & SEND. Check fitness of SEND data against the specifications
- Maintain “**single-truth**” of collected data in an accessible data store for consistency between study reports and SEND Datasets
- Represent **reportable data** in a repository:
 - ✓ Data for tabulation or analysis for Study Reports in dose groups
 - ✓ Data for columnar representation in SEND Trial Design and Trial Sets using CDISC terminology
 - ✓ Final SEND datasets for delivery to sponsors, or by sponsors for submission to the FDA
- Apply **data QA tools/processes** for final release of submission ready SEND packages

Planning for SEND at Study Start

- **Optimize** protocol template to capture SEND required variables
 - Example: Include “Sponsor Identifier” and “Study Monitor” as fields to be populated in the study protocols
- **Harmonize** terms from Controlled Terminology (CT) lists in study protocol template and LIMS systems
 - Example: Report study type using the same terms from the CDISC CT List
- **Uniformity** in data collection. Ensure all data are collected in the same format.
 - Makes automation of SEND datasets across different data types possible
- **Specify** your SEND dataset during study planning. Ensures consistency in naming conventions across disparate data sources.

Using the Study Protocol to Specify SEND Domains

15.3 Observations of Animals Starting on Study Day 1

Animals will be observed and data will be recorded as indicated in the following table.

Table 6: Observations of Animals Starting on Day 1

Procedure
Cageside Observations (mortality, moribundity, and general health)
Physical Examinations (skin and fur characteristics, eye and mucous membranes, respiratory, circulatory, autonomic and central nervous systems, somatomotor and behavior patterns, and injection sites)
Dermal Observations (Draize Scoring)
Body Weights
Food Consumption - Quantitative
Ophthalmologic Examinations
Body Temperature

Corresponding SEND Domain



Clinical Observations



Clinical Observations



Clinical Observations



Body Weights



Food Consumption



Clinical Observations



Vital Signs

Specify SEND Domains - example

Study Data Definition for 2275-13304 (v)

Dataset Level | Variable Level | Value Level Metadata | Codelist and Controlterms | External Dictionaries | Computational Algorithm | Support

Exclude | Include | Save | Clear

☒	*Domain	*Dataset Name	*Description	SAS Dataset Name	Dataset File Name	Status	Sources
☒	BG	BG	Body Weight Gains	BG	bg.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	BW	BW	Body Weights	BW	bw.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	CL	CL	Clinical Observations	CL	cl.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	CO	CO	Comments	CO	co.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	DD	DD	Death Diagnosis	DD	dd.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	DM	DM	Demographics	DM	dm.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	DS	DS	Disposition	DS	ds.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	EG	EG	ECG Test Results	EG	eg.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	EX	EX					SEND IG 3.0 to NSDR Import...
☐	FW	FW					SEND IG 3.0 to NSDR Import...
☐	LB	LB					SEND IG 3.0 to NSDR Import...
☐	MA	MA					SEND IG 3.0 to NSDR Import...
☐	MI	MI					SEND IG 3.0 to NSDR Import...
☐	OM	OM	Organ measurements	OM	om.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	PC	PC	Pharmacokinetics Concentr...	PC	pc.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	PM	PM	Palpable Masses	PM	pm.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	POOL...	POOLDEF	Pool Definition	POOLDEF	pooldef.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	PP	PP	Pharmacokinetics Parameters	PP	pp.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	RELREC	RELREC	Related Records	RELREC	relrec.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	SC	SC	Subject Characteristics	SC	sc.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	SE	SE	Subject Elements	SE	se.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	TA	TA	Trial Arms	TA	ta.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	TE	TE	Trial Elements	TE	te.xpt	Include	SEND IG 3.0 to NSDR Import...
☒	TF	TF	Tumor Findings	TF	tf.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	TS	TS	Trial Summary	TS	ts.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	TX	TX	Trial Sets	TX	tx.xpt	Include	SEND IG 3.0 to NSDR Import...
☐	VS	VS	Vital Signs	VS	vs.xpt	Include	SEND IG 3.0 to NSDR Import...

Exclude the selected datasets from:

☒ Study

☒ SEND IG 3.0 to NSDR Import Mapper

Ok Cancel

Using Study Design to build the SEND Trial Design

15.1 Group Designation and Dosage Levels

Animals will be assigned to groups as noted in the table below.

Table 5: Group Designation and Dosage Levels

Group	Treatment	Test Article Dosage (mg/kg)	Number of Animals			
			Main Phase		2-Week Recovery Phase	
			Males	Females	Males	Females
1	Control Article	0	5	5	5	5
2	Test Article	5	5	5	5	5
3	Test Article	10	5	5	5	5
4	Test Article	50	5	5	5	5

Trial Sets

1NR	Vehicle Control once daily, Nonrecovery
2NR	5 mg/kg once daily, Nonrecovery
3NR	10 mg/kg once daily, Nonrecovery
4NR	50 mg/kg once daily, Nonrecovery
1R	Vehicle Control once daily, Recovery
2R	5 mg/kg once daily, Recovery
3R	10 mg/kg once daily, Recovery
4R	50 mg/kg once daily, Recovery

Treatment Elements

TRT01	Vehicle Control
TRT02	5 mg/kg Drug, once daily
TRT03	10 mg/kg Drug, once daily
TRT04	50 mg/kg Drug, once daily
RECO	Recovery

Trial Arms Order of Elements

1	TRT01
2	TRT02
3	TRT03
4	TRT04
1R	TRT01, RECO
2R	TRT02, RECO
3R	TRT03, RECO
4R	TRT04, RECO

Specifying Trial Design Domains

Codelist Name						Short Codelist Name				☐	*Submission Value	Preferred Term	Submission...	Preferred S...	Rank	Ord...	Code	Ext...
☐	*Name	*Short Name	C...	SAS ...	D...	Name	Short...	Co...	S...	☐	Vehicle Control							
☐	Element Co...	ETCD			text					☐	5 mg/kg Drug, once daily							
☐	Element	ELEMENT			t...					☐	10 mg/kg Drug, once daily							
☐	 Laboratory...	LBSPEC			text					☐	50 mg/kg Drug, once daily							
☐	 Clinical Pat...	LBTEST			text					☐	Recovery							

Study Data Definition for 2275-13304 (v)

Dataset Level

Variable Level

Value Level Metadata

Codelist and Controlterms

External Dictionaries

Computational Algorithm

Support Documents

Exclude

Include

Save

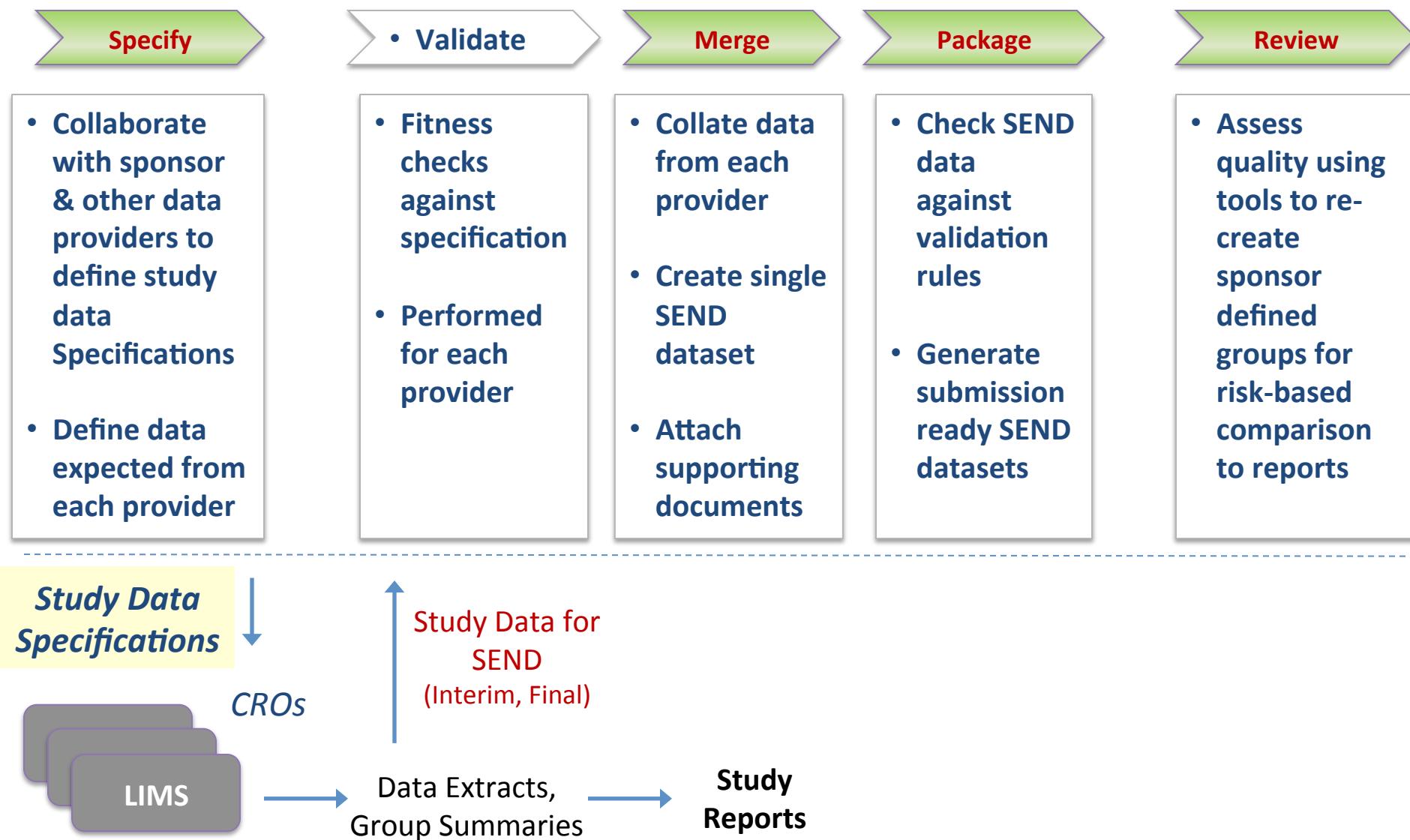
Clear

☐	*Domain	*Variable Name	Description	Value Coded By	Codelist or External Codelist or Format	Role	SAS Field Name
☐	TE	STUDYID	Study Identifier			Identifier	STUDYID
☐	TE	DOMAIN	Domain Abbreviation			Identifier	DOMAIN
☐	TE	ETCD	Element Code			Topic	ETCD
☐	TE	ELEMENT	Description of Element	Codelist	Element 	Synonym Qualifier	ELEMENT
☐	TE	TESTRL	Rule for Start of Element			Rule	TESTRL
☐	TE	TEENRL	Rule for End of Element			Rule	TEENRL
☐	TE	TEDUR	Planned Duration of Element			Timing	TEDUR

Specifying Controlled and Preferred terms for SEND

Dataset Level		Variable Level		Value Level Metadata		Codelist and Controlterms		External Dictionaries		Computational Algorithm		Support Documents																																																																																															
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A global SEND process for CROs and Sponsors



Getting SEND done right

- ***“Fit for review”*** SEND datasets for FDA submissions must be complete, consistent and technically compliant
- Don’t run two separate processes for SEND and study reports to avoid *data quality, increase downstream costs and delay FDA reviews* once submitted
- High quality SEND datasets with reduced costs can result from:
 - ✓ Establishing data specifications and ensuring that the right data are collected (***“Plan the Work”***)
 - ✓ Using a single truth data resource for study reporting and automation of SEND data assembly (***“Work the Plan”***)
 - ✓ Applying tools to easily compare reports and SEND datasets (***“Review the Work Product”***)

Perspectives on SEND from a CRO

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Executive Vice President
Smithers Avanza Toxicology Services

Starting your SEND Journey?

- Assign a point person with appropriate responsibility, accountability and operational knowledge
- Evaluate QA/QC processes
 - **QAU must be engaged with your operational processes**
 - Data correction procedures must address source data, not just report output
- Determine SEND Strategy
 - SEND internal ownership
 - Timing?
 - **SEND As A Service**
 - Selection of a qualified partner

Plan for SEND at Study Start

15.3 Observations of Animals Starting on Study Day 1

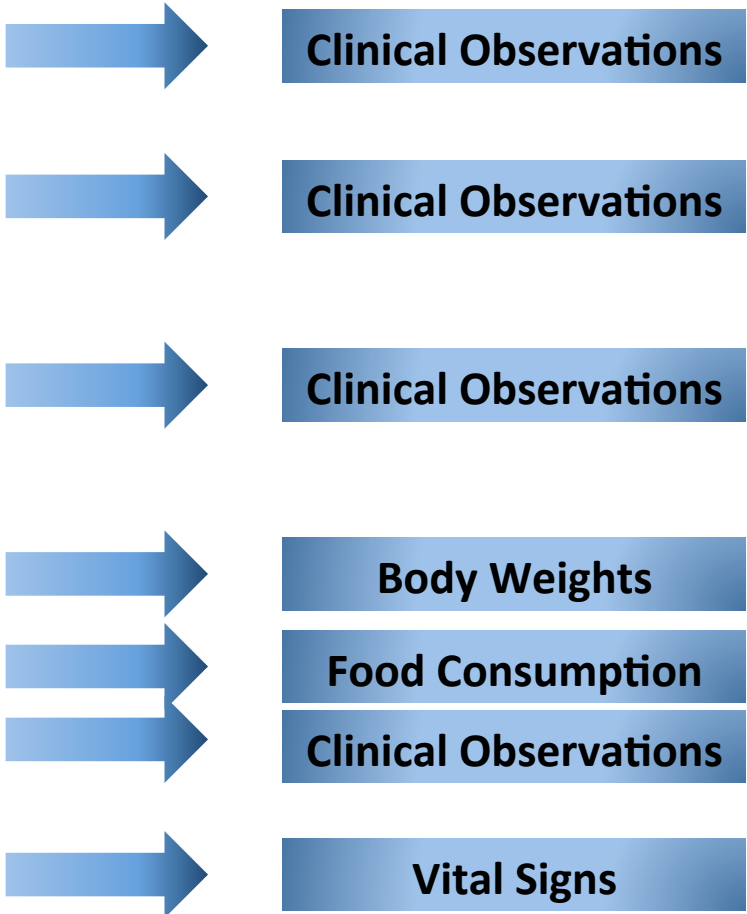
Animals will be observed and data will be recorded as indicated in the following table.

Table 6: Observations of Animals Starting on Day 1

Procedure
Cageside Observations (mortality, moribundity, and general health)
Physical Examinations (skin and fur characteristics, eye and mucous membranes, respiratory, circulatory, autonomic and central nervous systems, somatomotor and behavior patterns, and injection sites)
Dermal Observations (Draize Scoring)
Body Weights
Food Consumption - Quantitative
Ophthalmologic Examinations
Body Temperature

Use the Study Protocol to Specify SEND Domains

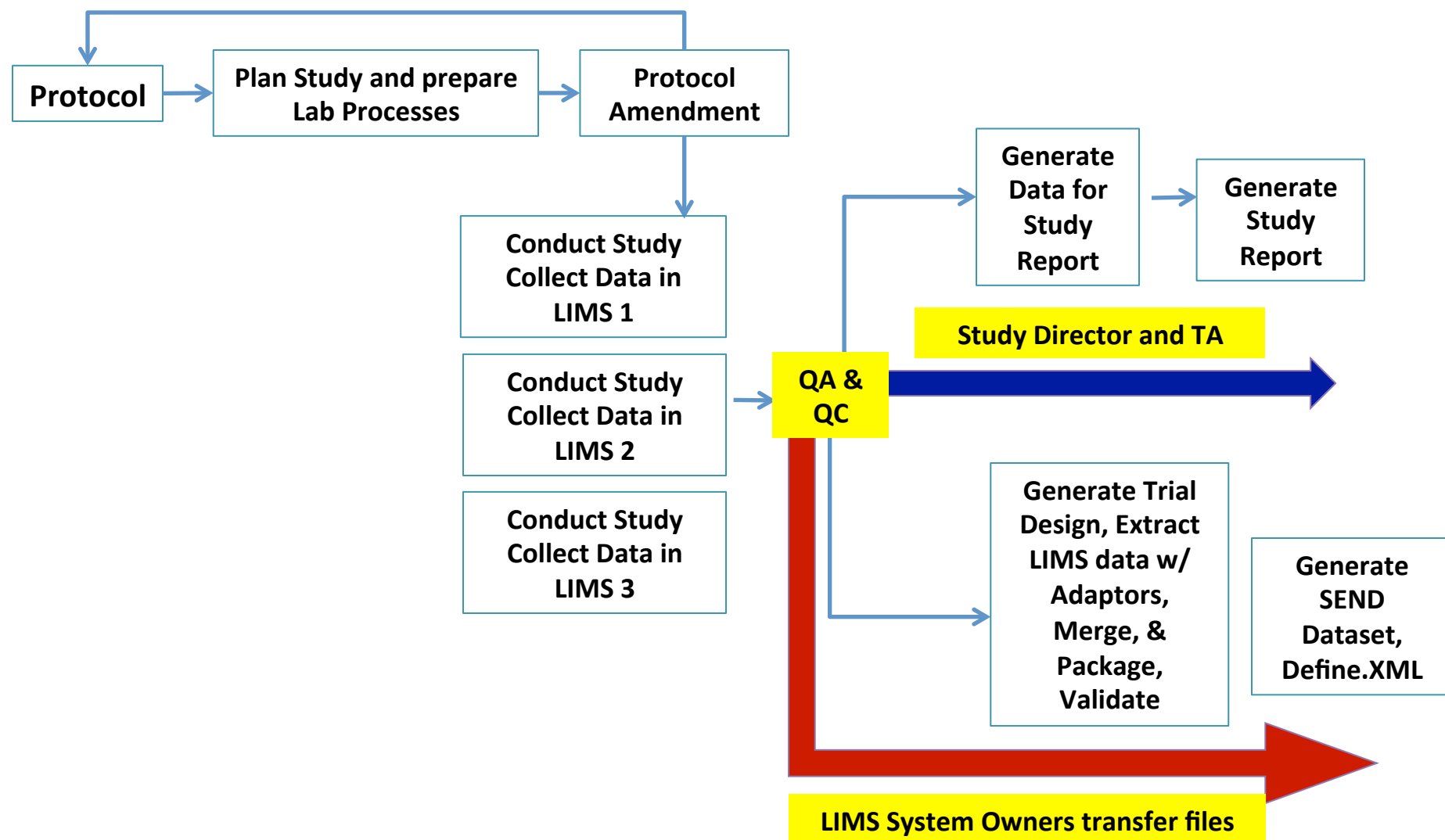
Corresponding SEND Domain



Maximize the Use of your LIMS Systems

- Current laboratory processes were not designed with SEND in mind: “***Optimization, Uniformity, Harmonization***”
 - Information necessary for preparation of SEND data sets is not necessary to set up studies in your system (*Optimize*)
 - Example: Include “Sponsor Identifier” and “Study Monitor” as fields to be populated in the LIMS system
- Maximize use of electronic data collection (*Uniformity*)
- Utilize Controlled Terminology (CT) lists in protocol template and LIMS (*Harmonize*)
 - Establish hierarchy for proper SEND mapping

QA/QC Process



SEND Impacts Everything!

Plan Now!